



Introduction of Agena Bioscience's

MassARRAY Overview

20 March, 2025

Genomic Market

Global Genomics Market

PRESCIENT & STRATEGIC
INTELLIGENCE
Where knowledge inspires strategy

North America
Largest Market
By Region (2019)

APAC
Fastest-Growing Market
By Region (2020-2030)

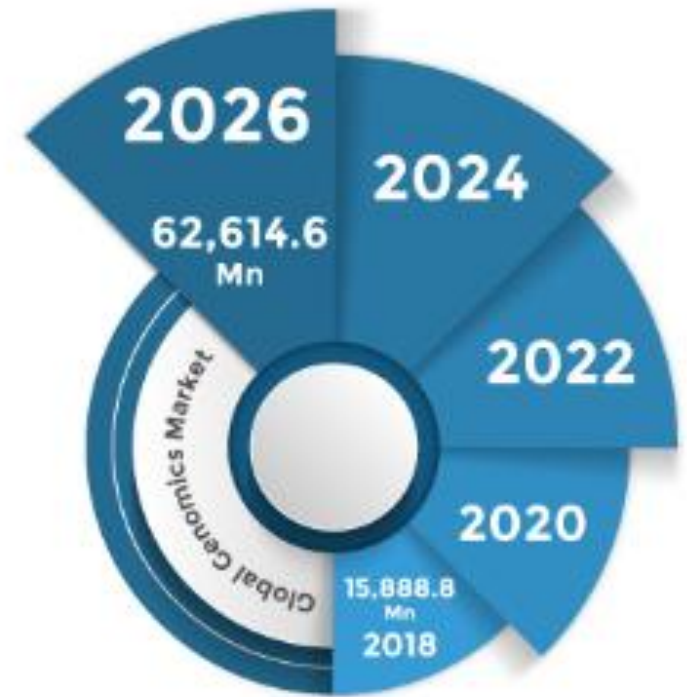


2019
Market Size
\$18.2
billion

2030
Market Size
\$68.0
billion

Market
Growth Rate
(2020-2030)
12.7%

Global Genomics Market (US\$ Mn), 2018 to 2026



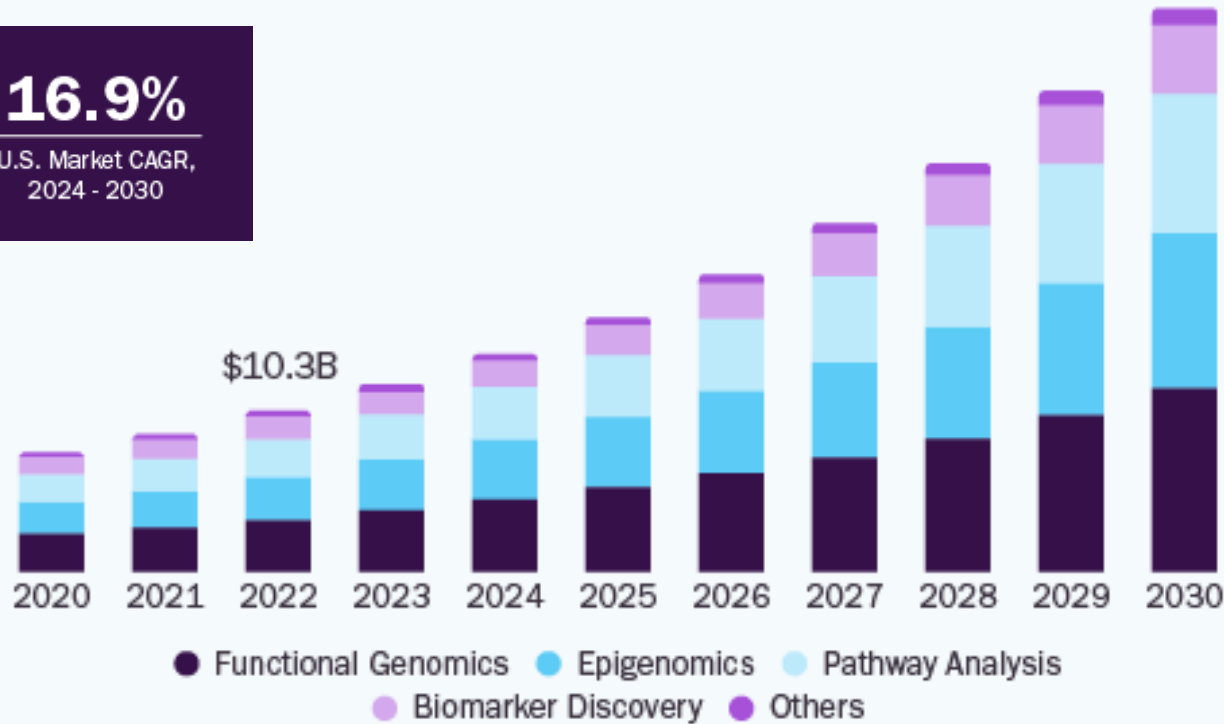
Genomic Market

U.S. Genomics Market

Size, by Application & Technology, 2020 - 2030 (USD Billion)

16.9%

U.S. Market CAGR,
2024 - 2030

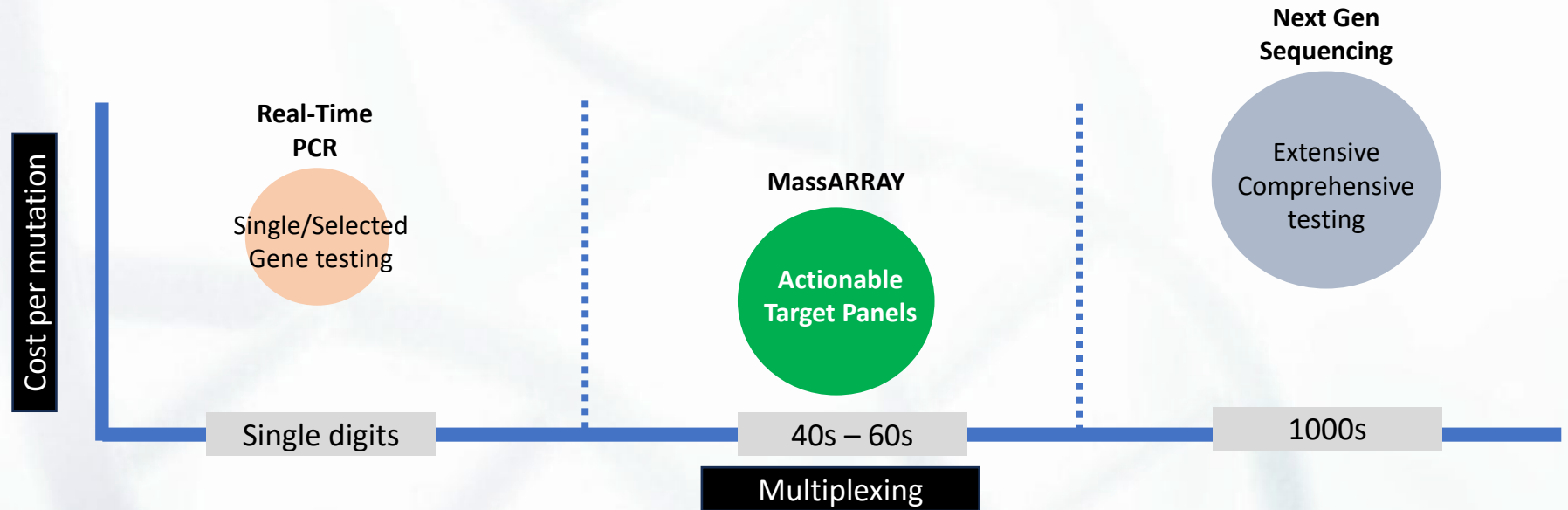
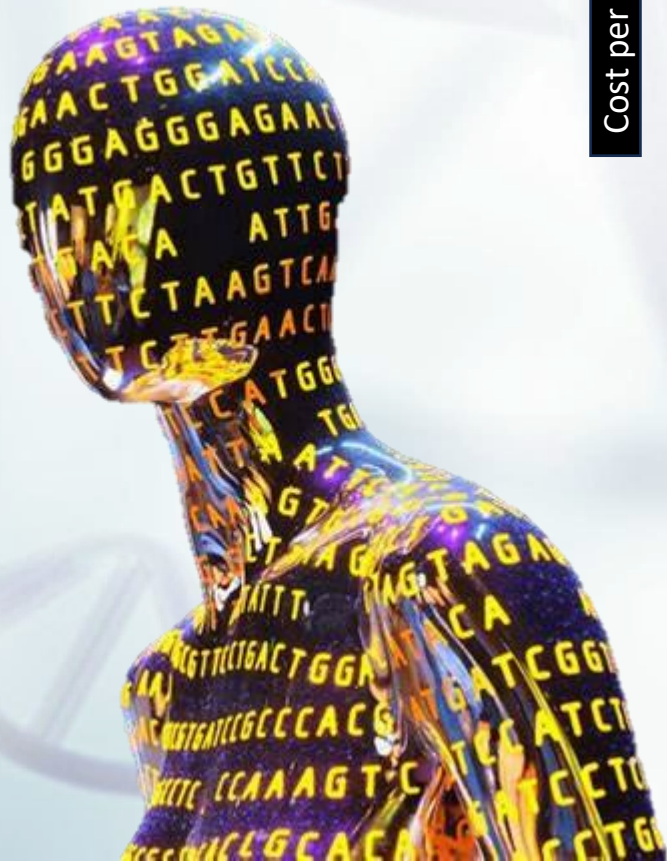


Global Precision Medicine Market–Analysis & Forecast 2017 – 2026



Market Segmentation

Robust, Flexible, and High Throughput



- De-Novo
- Single SNPs

- De-Novo
- Multiplexing
- Local validation
- Local Primer Production
- LDT develops test – Patent
- Cost efficiency due to multiplexing SNPs per well

- Novo
- Comprehensive testing
- Import from Principle
- BioIT requirements

Kockum, I., Huang, J., & Stridh, P. (2023). Overview of Genotyping Technologies and Methods. *Current Protocols*, 3(4), e727.



Overview of Genotyping Technologies and Methods

Ingrid Kockum,¹ Jesse Huang,¹ and Pernilla Stridh^{1,2}

¹Center for Molecular Medicine, Department of Clinical Neuroscience, Karolinska Institutet, Stockholm, Sweden

²Corresponding author: pernila.stridh@ki.se

Published in the Essential Lab Techniques section

Genetics is a cornerstone of molecular biology, and there have been significant developments in genotyping technologies during the last decades. Genotyping can be used for a wide range of applications, such as genealogy, assessing risks and causes for common diseases and health conditions, animal and human research, and forensic investigations. So how do you perform a genetic test? This overview covers key concepts in genetics, the development of genotyping methods, and a comparison of several techniques, including PCR, microarrays, and sequencing. A general process of the steps involved in genotyping, from DNA preparation to quality control, is described with relevant protocols referenced. Different types of DNA variants are illustrated, including mutations, SNP, insertions, deletions, microsatellites, and copy number variations, with examples of their involvement in disease. We discuss the utilities of genotyping, such as medical genetics, genome-wide association studies (GWAS), and forensic science. We also provide tips for quality control, analysis, and results interpretation to help the reader design and perform a genetic study or scrutinize such studies from the literature. © 2023 The Authors. *Current Protocols* published by Wiley Periodicals LLC.

Keywords: genetics • genotyping • GWAS • methodology • microarray • NGS • PCR

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INTRODUCTION

Genetics is a keystone of molecular biology

vide a foundation for academics (undergraduates/graduates) and industry professionals.

Table 2 Overview of Genotyping Technologies

Name	Cost	# markers	Pro	Con	Ref.
PCR-RFLP	+	1	Easy to run in any lab, fast, flexible	Time consuming, manual inspection	(Saiki et al., 1985)
Allele-specific PCR	+	1	Easy to run in any lab, fast, flexible	Time consuming, manual inspection	(Gaudet et al., 2009)
TaqMan PCR	+	1	Standardized, more accurate	Manual inspection, requires specific equipment	(Hui et al., 2008)
Microsatellite	+	1	Robust, do not require specific equipment, flexible	Low resolution, manual inspection, time consuming	(Weber & May, 1989)
Pyrosequencing	++	1	Captures all potential alleles	Time consuming, manual inspection, requires specific equipment	(Kreutz et al., 2013)
iPLEX	++	60 ++	Multiplex assay	Manual inspection, requires specific equipment	(Gabriel et al., 2009; Tang et al., 1999)
Multiplex TaqMan	++	up to 100	Multiplex assay	Requires specific equipment	(Martínez-Cruz et al., 2011)
Genotyping arrays	+++	50k-2 mil	Multiplex assay	Requires specific equipment and expertise	(Verlouw et al., 2021)
NGS-Exome	++++	25000	captures all coding variants	Requires specific equipment and expertise, demanding processing	(Seaby et al., 2016)
NGS-Whole Genome	+++++	up to 40 mil	captures all variants	Requires specific equipment and expertise, demanding processing	(Slatko et al., 2018)

Rough relative cost per genotype is indicated by +, since prices vary over time.

Agena Bioscience®

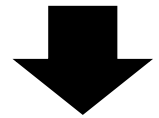


- Headquarters in San Diego, California, USA
- Agena markets our products in over 30 countries worldwide through direct sales offices in Germany, China and Australia,

SEQUENOM®



(BEFORE 2014)



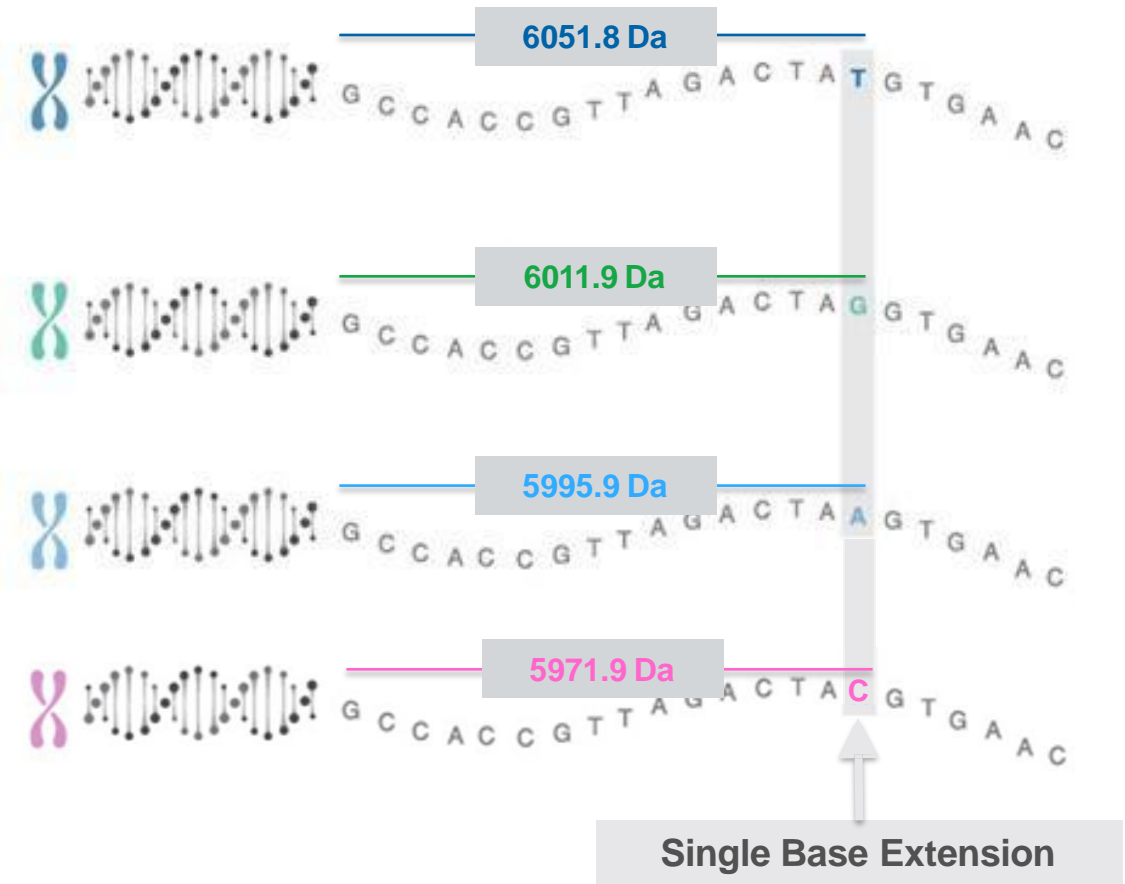
Agena
BIOSCIENCE



MassARRAY system

Mass-Detection With The MassARRAY

Single Nucleotide Polymorphism (SNPs)

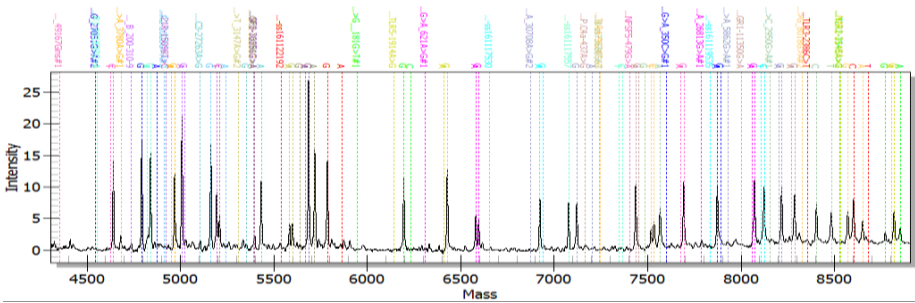


The MassARRAY is unique technology to detect DNA mutation SNP, Insertion, Deletion

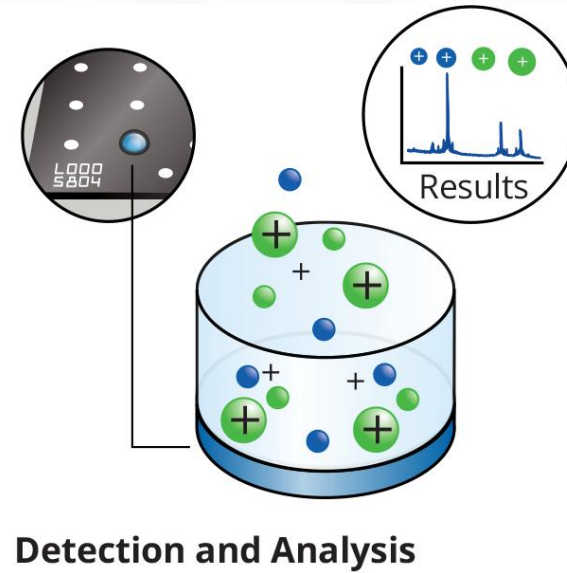
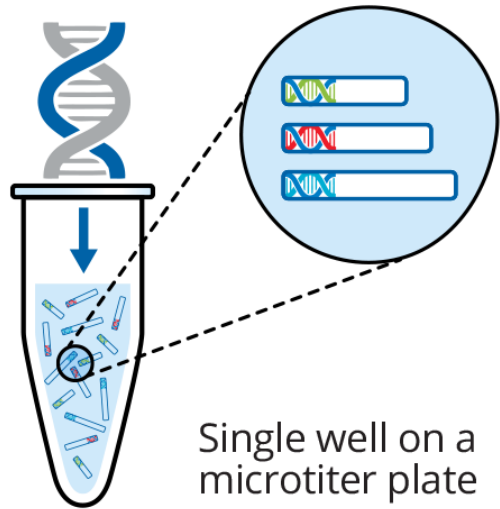
Dalton
Unit of Mass

The Dalton (Da) is a unit of mass widely used in physics and chemistry.

Cytosine (C)	Adenine (A)	Guanine (G)	Thymine (T)
247.2 Da	271.2 Da	287.2 Da	327.1 Da



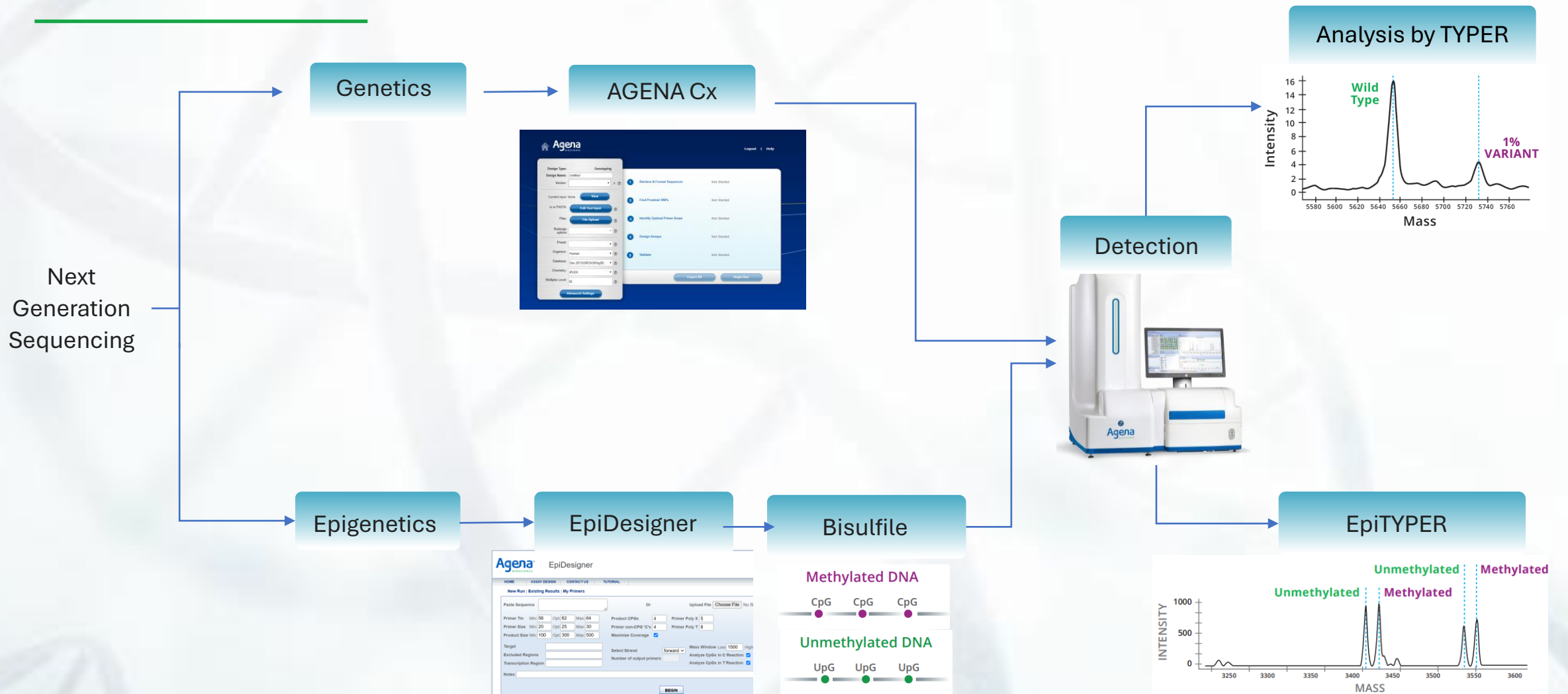
MassARRAY Technology



**Multiplex
PCR**

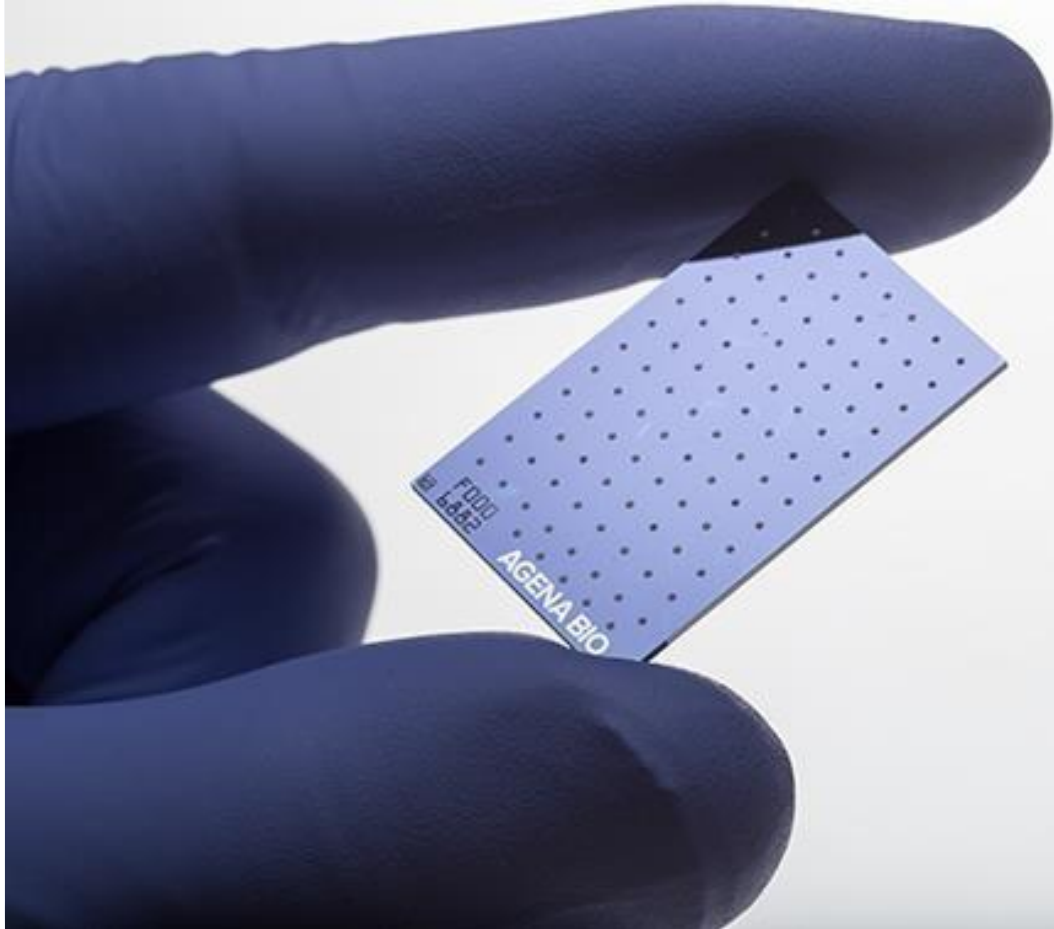
**MALDI-TOF
Mass Spectrometry**

MassARRAY



SpectroCHIP Arrays

A Powerful, Open Source Array for Custom Multiplex Assay Designs



Composed of 96 or 384 inert pads evenly spaced across a matrix.

- Each pad binds to any DNA analyte mixture
- Enables side-by-side testing of different assays
- Utilizes nanoliters of sample

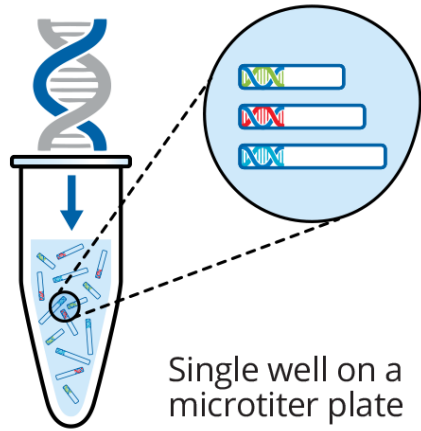
Genotype up to 50* (usually around 30) genetic variants on each SpectroCHIP Array pad

Flexible biomarker detection.

■ SNPs ■ Insertions ■ Deletions ■ Translocations ■ CNV

*actual number may vary depending on chemistry and application

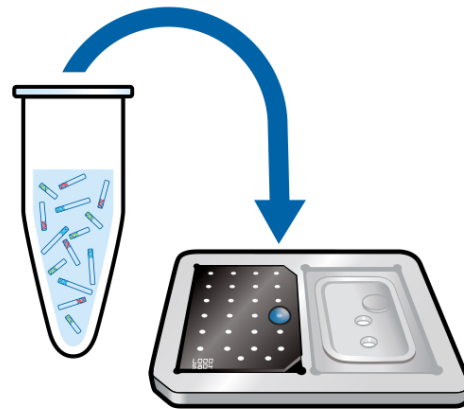
SAMPLE PROCESS JOURNEY



Single well on a microtiter plate

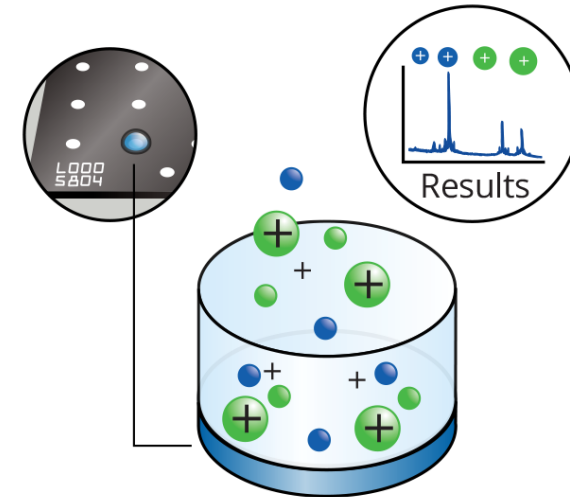
Endpoint PCR

Amplify and extend up to 40 target-specific DNA fragments in a single reaction.



Transfer Analyte

Transfer a small amount of sample to a single pad on the SpectroCHIP® Array.

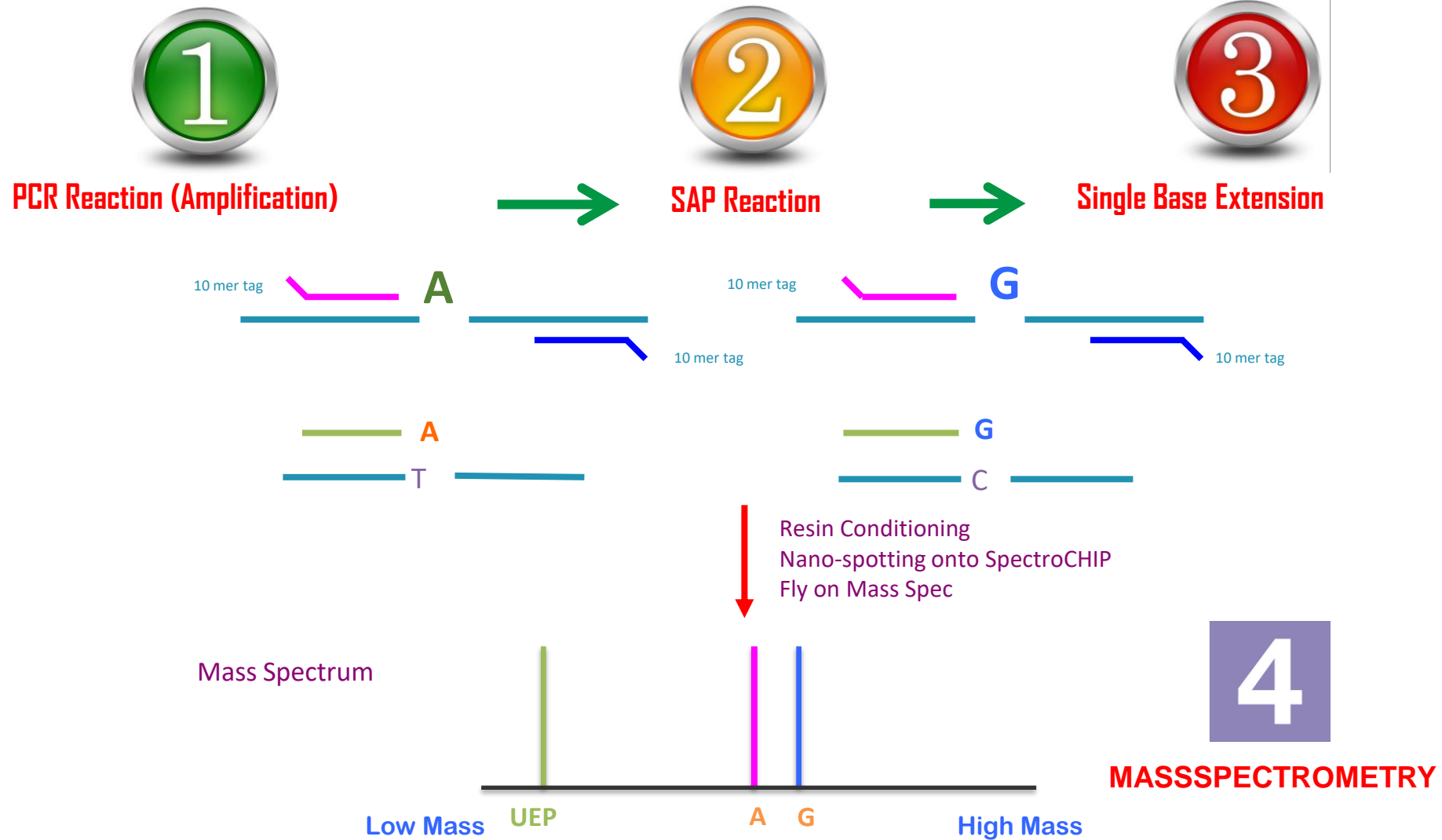


Detection and Analysis

Multiple tests can be run on a single SpectroCHIP Array. Hundreds of mutations can be tested per sample.

* Use multiple reactions for >40 targets if required.

GENOTYPING REACTION : SNP (A/G)

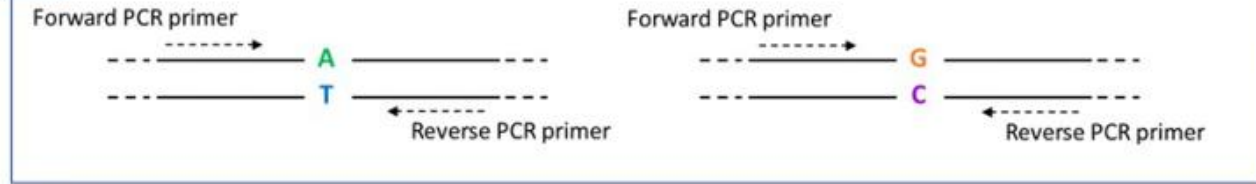


PCR : Polymerase Chain Reaction

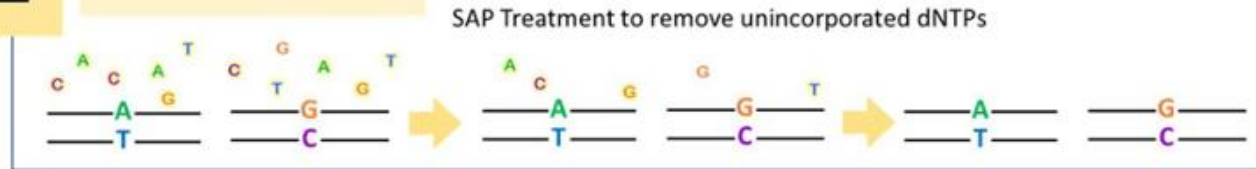
SAP : Shrimp Alkaline Phosphatase

UEP : Unextension Primer

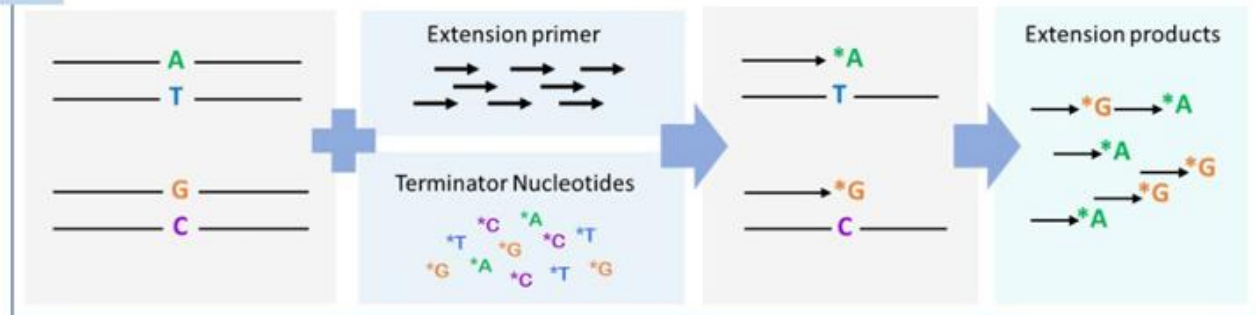
1 PCR Reaction



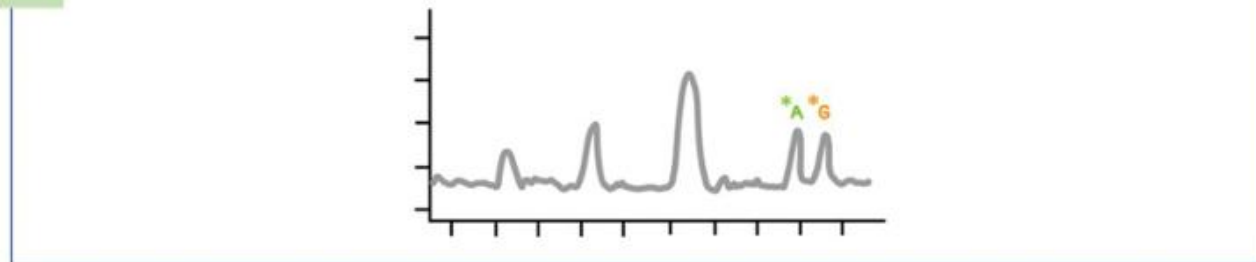
2 SAP Reaction



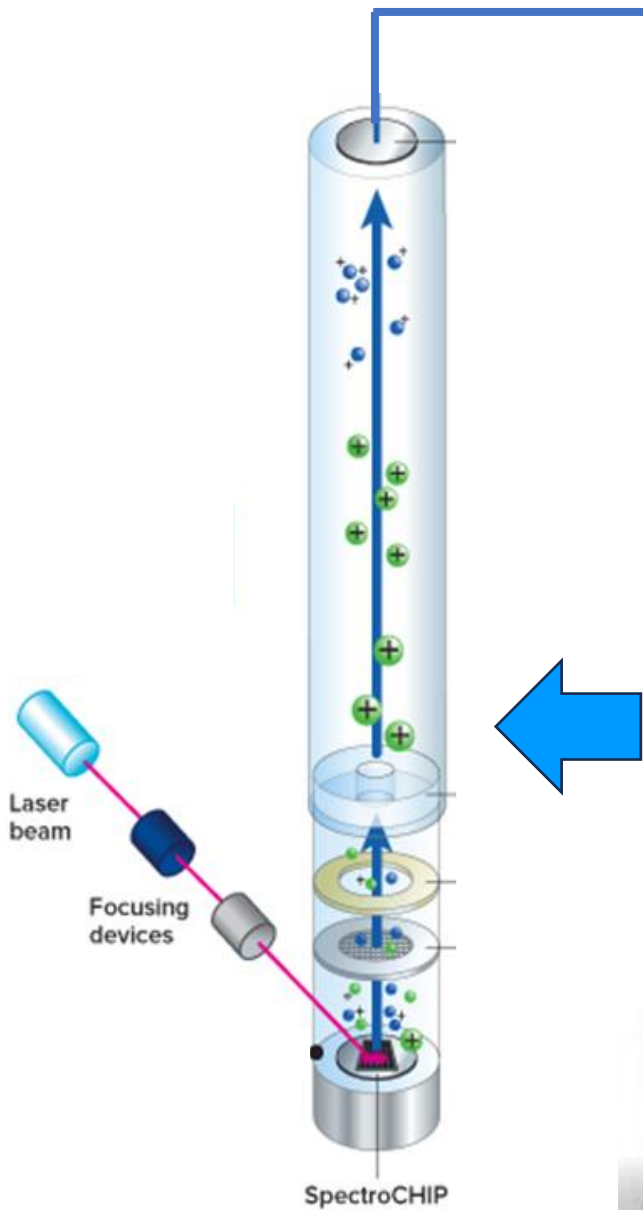
3 Single Base Extension



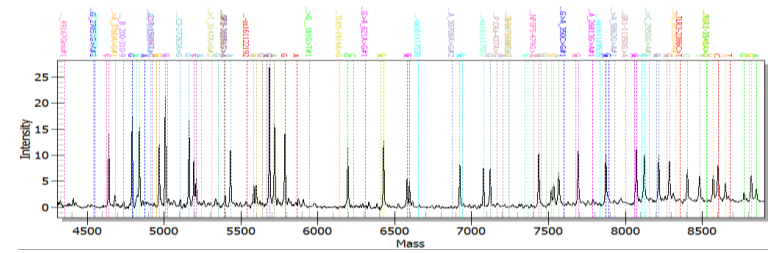
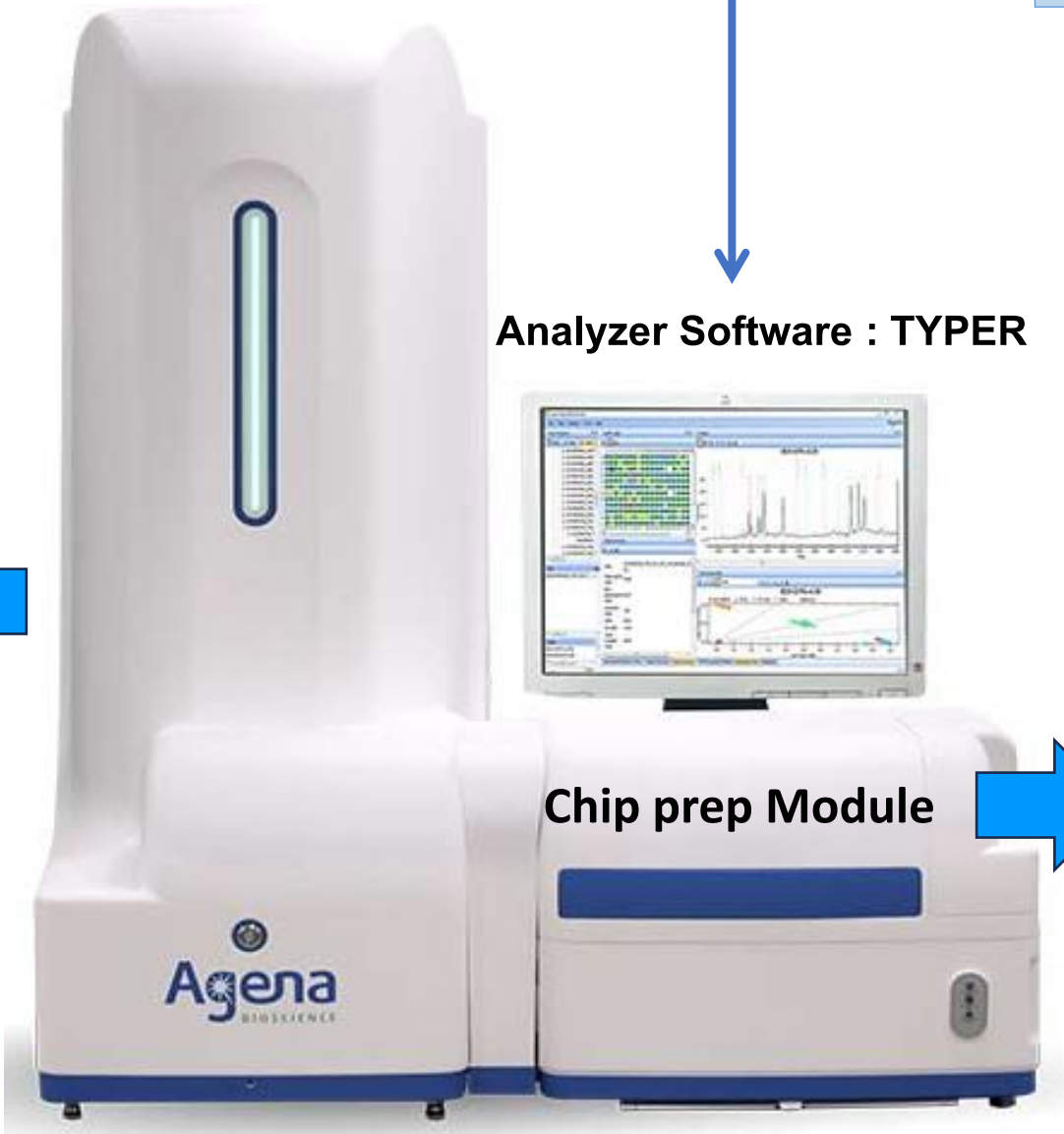
4 MASS SPECTROMETRY



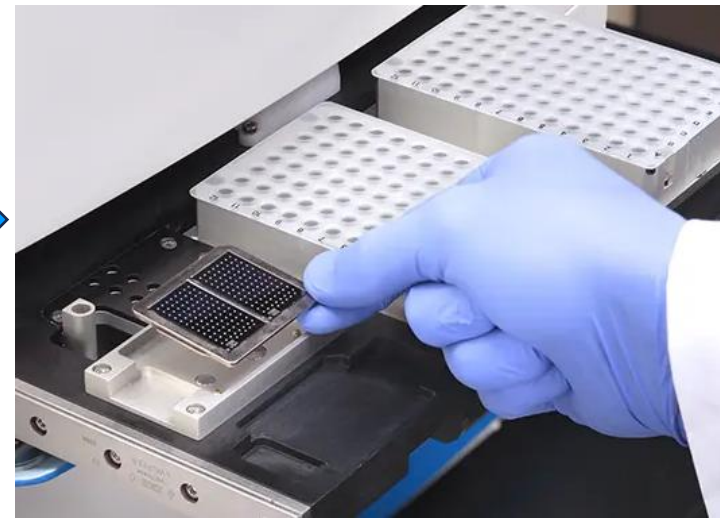
MassARRAY platform



Mass Analyzer



Platform: 96 wells x 2 plates



PATENTED "SPECTROCHIP"

REPORT: EASE INTERPRETATION

TyperAnalyzer

File View Options Tools Help

Project Explorer

Customer/Project/Plate/Experiment/Chip

Chip Experiment

F0028268_RECALL_869708... 3

Assay

CYP2D6_Exon1-3
CYP2D6_Exon9-1
CYP2D6_Exon9-2
CYP2D6_Exon9-3
CYP2D6_Intron2-1
CYP2D6_Intron2-2
CYP2D6_Intron2-4
CYP2D6_Intron4-1
CYP2D6_Intron4-2
CYP2D6_Intron4-3
CYP2D6_Intron6-1
CYP2D6_Intron6-2
CYP2D6_Intron6-3
CYP2D6_Intron7-1
CYP2D6_Intron7-3
CYP2D6_Intron7-4

Traffic Light

	1	2	3	4	5	6	7	8	9	10	11	12
A	●	●	●	●	●	●	●	●	●	●	●	●
B	●	●	●	●	●	●	●	●	●	●	●	●
C	●	●	●	●	●	●	●	●	●	●	●	●
D	●	●	●	●	●	●	●	●	●	●	●	●
E	●	●	●	●	●	●	●	●	●	●	●	●
F	●	●	●	●	●	●	●	●	●	●	●	●
G	●	●	●	●	●	●	●	●	●	●	●	●
H	●	●	●	●	●	●	●	●	●	●	●	●

Chip Summary

Chip F0028268_RECALL_869708_(3) (3)

Conservative Calls 612
Moderate Calls 615
Aggressive Calls 287
User Calls 0
Calls 1514
No Calls 598
Total Possible Calls 2112
Call Rate 71.7
Negative Controls 0
Negative Control Calls 0
Neg. Control Call Rate 0.0
Positive Controls 0
Positive Control Calls 0
Pos. Control Call Rate 0.0
Assays 22

Details

5403.043, 12.478

CYP2D6_Intron7-4

Well: D08 Sample: 5

Assay	Call	Description
YP2D6_5Pri...	T	A.Conserv
YP2D6_5Pri...	T	A.Conserv
YP2D6_5Pri...	C	A.Conserv
YP2D6_5Pri...	C	A.Conserv
YP2D6_Exon...	G	B.Moderat
YP2D6_Exon...	A	A.Conserv
YP2D6_Exon...	A	A.Conserv
YP2D6_Exon...	G	A.Conserv
YP2D6_Exon...	CA	A.Conserv
YP2D6_Exon...	A	A.Conserv
YP2D6_Intro...	G	A.Conserv
YP2D6_Intro...	A	A.Conserv
YP2D6_Intro...	C	A.Conserv
YP2D6_Intro...	A	A.Conserv
YP2D6_Intro...	A	A.Conserv
YP2D6_Intro...	T	A.Conserv
YP2D6_Intro...	A	A.Conserv
YP2D6_Intro...	C	A.Conserv

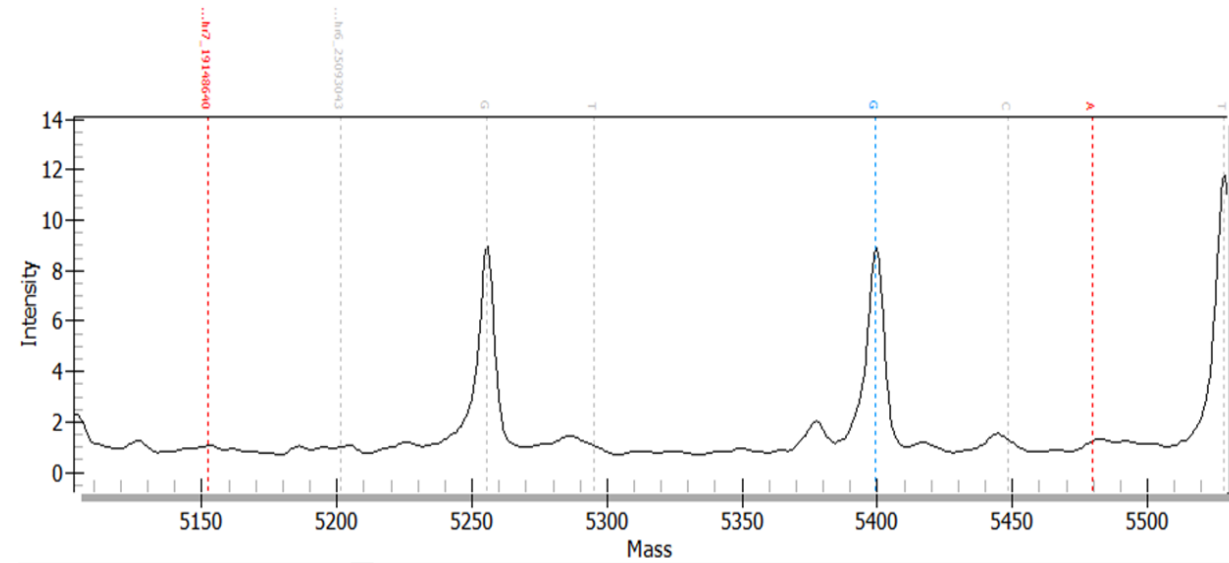
Expected Peaks	Name	Sequence	Mass
Probe	UEP.CYP2D6...	AAAAACCCCCCGGGTT	541
Analyte	C	AAAAACCCCCCGGGTT	566
Analyte	A	AAAAACCCCCCGGGTT	568

Automated Data Error Checking Assay Summary Chip Summary Post Processing Clusters Call Cluster Plot Histogram Details

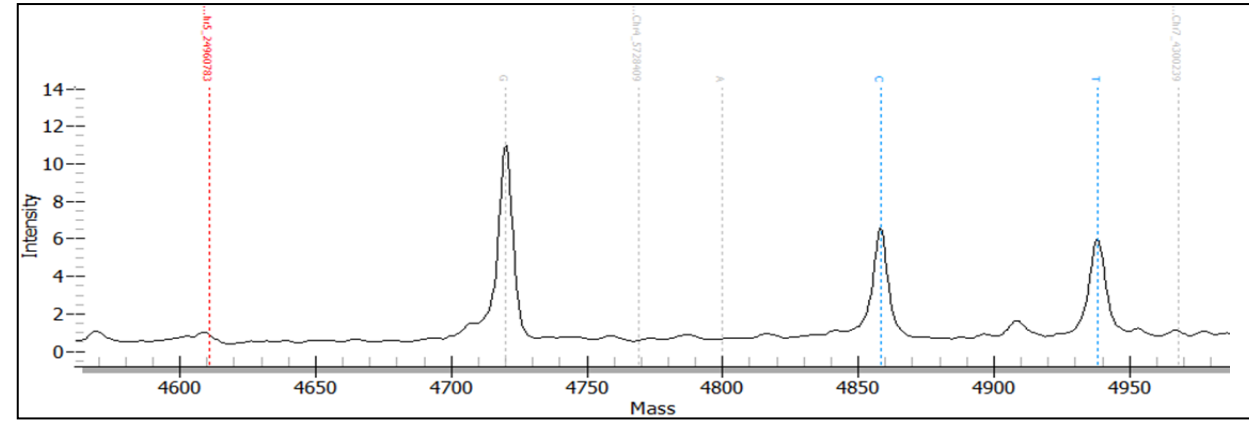
Done

SCRL

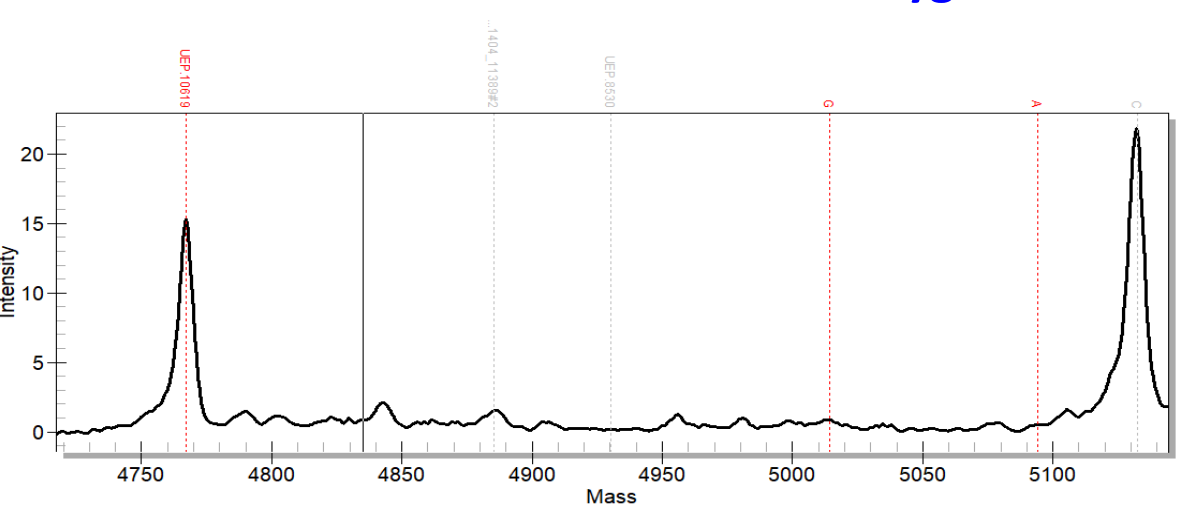
REPORT: Spectrum peak



Call: Homozygous GG

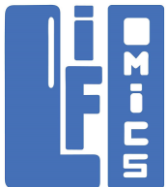


Call: Heterozygous CT

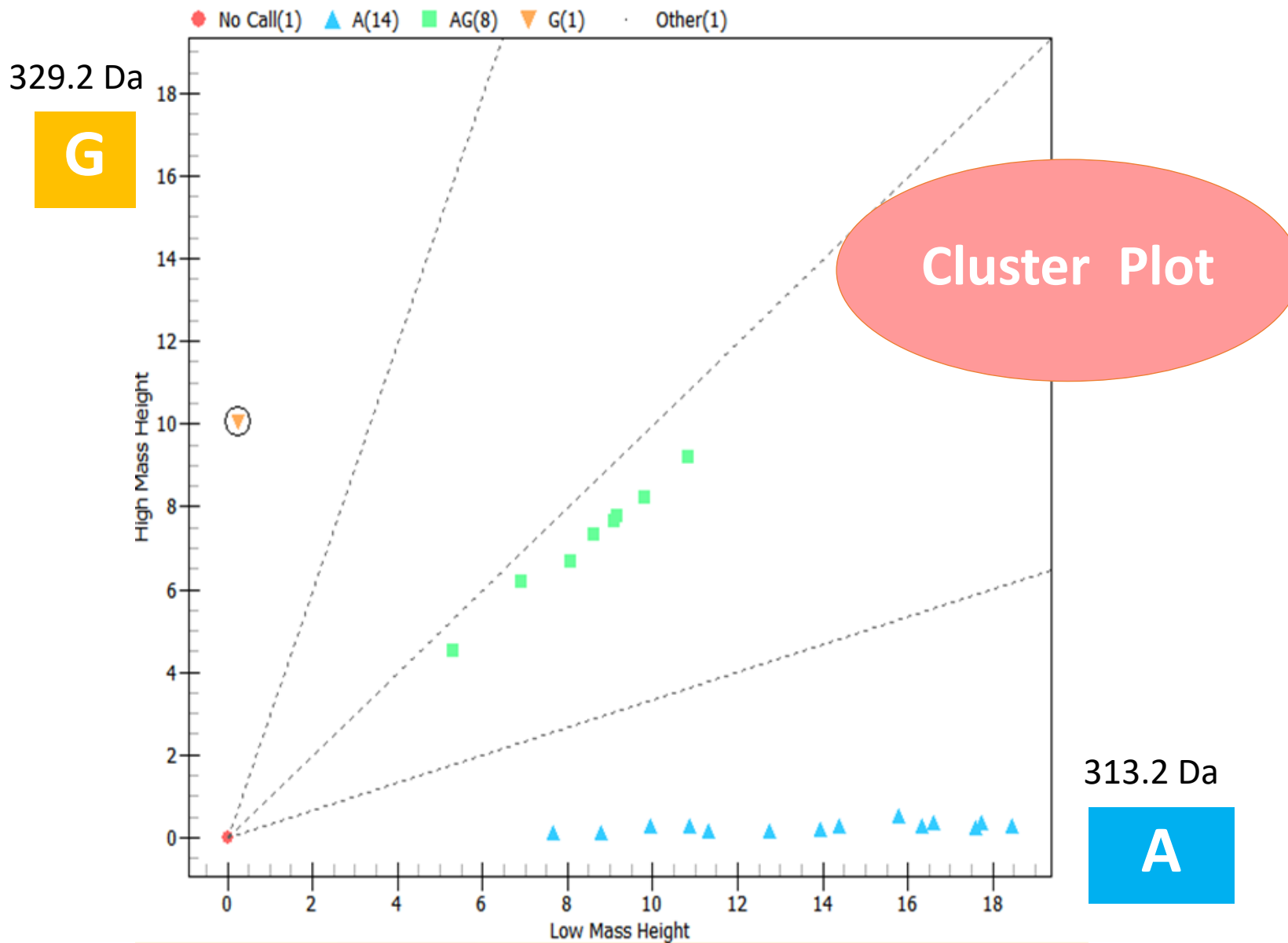


Only UEP Peak

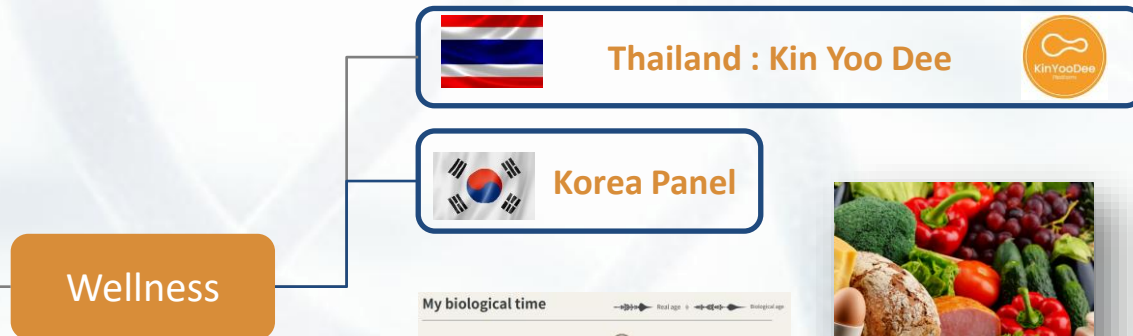
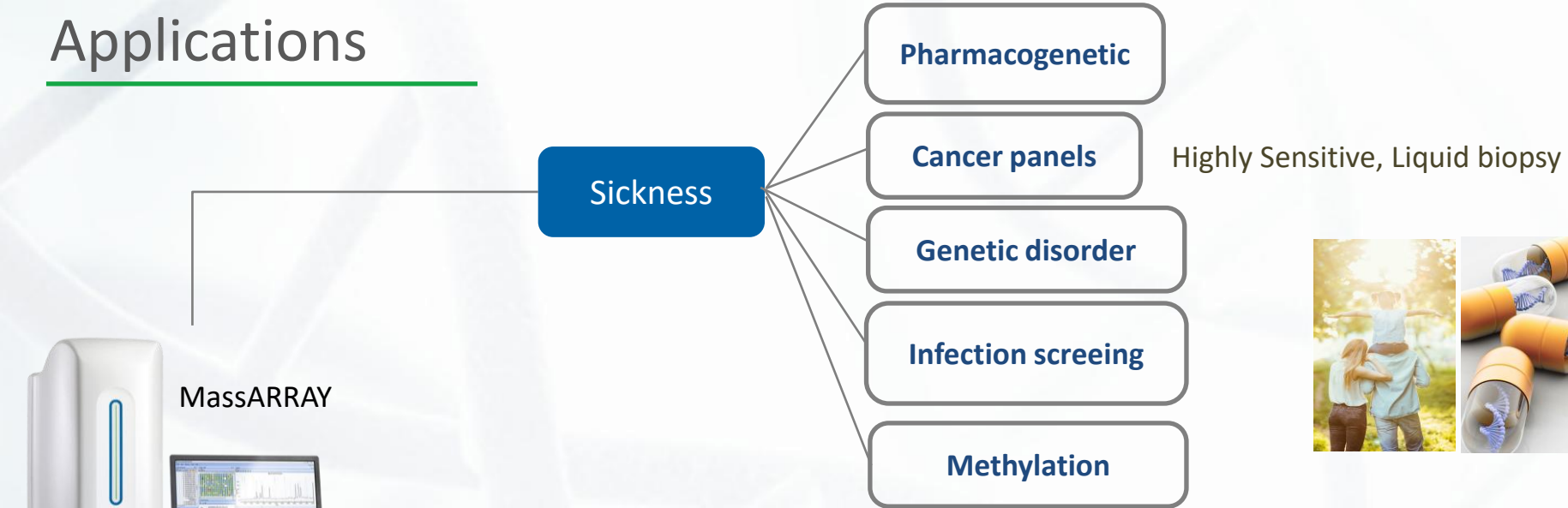
No Call



REPORT: EASE INTERPRETATION



Applications



SICKNESS

ONCOLOGY (CANCER)

- EMPHASIZE ADVANCEMENT
- PREDICTIVE FACTORS
- PROGNOSIS FACTORS
- TARGET THERAPIES SELECTION



Tissue specimen
Liquid Biopsy Panels

PHARMACOGENOMICS (PRECISION MED)

- SCAN ADME GENES
- PREDICT DRUG RESPONSE
- SIDE EFFECT, ADRs
- DRUG SURVEILLANCE FOR SENSITIVE PATIENTS



VeriDose[®] CYP2D6 CNV Panel
VeriDose[®] DPYD Panel

GENETIC DISORDERS

- ULTRASENSITIVE MATERNAL OR PRENATAL SCREENING
- LESS INVASION SCREENING



Hearing loss
Wilson's disease
Cystic Fibrosis Mutation

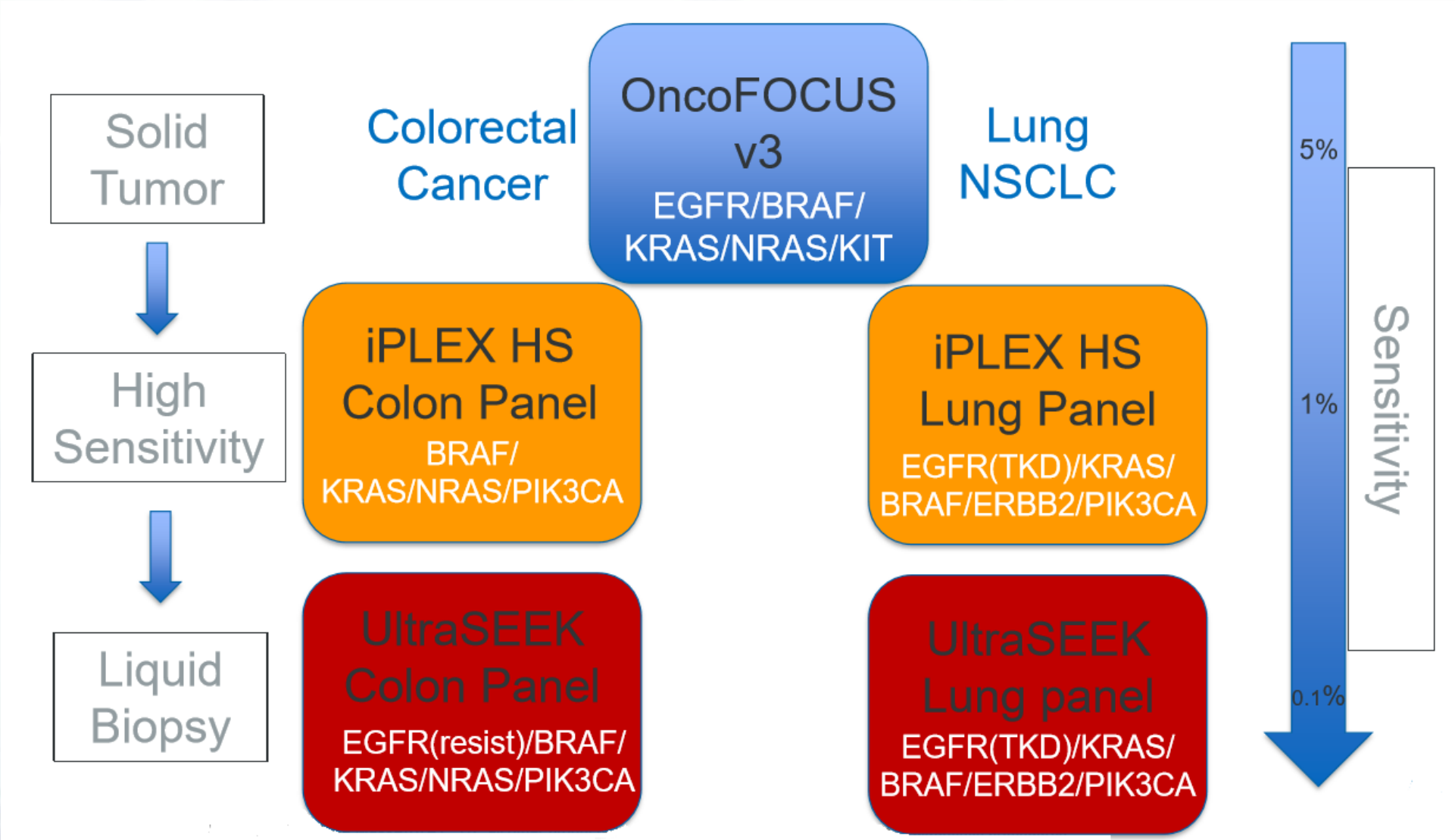
INFECTIOUS APPLICATIONS

- FAST SCREENING
- BACTERIA
- VIRUSES
- MYCOPLASMA
- NON CULTURE



Sexually transmitted disease
Human Papilloma Virus
Mycobacterium

MASSARRAY : Targeted Somatic Mutation Panels



PHARMACOGENETICS

VeriDose[®] CYP2D6 CNV Panel

CYP2D6 copy number variation (CNV)

CYP2D6/CYP2D7 “hybrid alleles” including exon 9 exchanges, *13 and *68.

22 points in 7 regions

VeriDose[®] Core Panel

85 SNPs/INDELS across 16 genes

Designed to robustly analyze low quality DNA

Genes Analyzed with the VeriDose Core Panel

• CYP2B6	• CYP3A4	• HLA-A	• TPMT
• CYP2C19	• CYP3A5	• HLA-B	• UGT1A1
• CYP2C9	• CYP4F2	• NUDT15	• VKORC1
• CYP2D6	• DPYD	• SLCO1B1	• 2C cluster

Patient Genetic Results

Gene	Genotype	Phenotype	Phenotype adjusted for concomitant medications*	Additional Comments
CYP2B6	*1/*1	normal	normal	Notes appear here.
CYP2C19	*1/*1	normal	poor	
CYP2C9	*1/*2	intermediate	intermediate	
CYP2D6	*1/*2	normal	normal	
CYP3A5	*3/*3	poor	poor	
HLA-A*31:01	-	positive	positive	
HLA-B*15:02	-	negative	negative	
HLA-B*57:01	-	negative	negative	
HLA-B*58:01	-	negative	negative	
NUDT15	*1/*1	normal	normal	
SLCO1B1	*1A/*1A	normal	normal	
TPMT	*1/*3A	intermediate	intermediate	
VKORC1	*1/*1	normal	normal	

Gene	Drugs
CYP2B6	Efavirenz
CYP2C19	Amitriptyline, citalopram, clobazam, clomipramine, clopidogrel, doxepin, escitalopram, esomeprazole, imipramine, lansoprazole, omeprazole, pantoprazole, prasugrel, sertraline, ticagrelor, trimipramine, voriconazole
CYP2C9	Aspirin, celecoxib, flurbiprofen, ibuprofen, lornoxicam, meloxicam, naproxen, phenytoin, piroxicam, tenoxicam, warfarin
CYP2D6	Amiodarone, amitriptyline, amphetamine, aripiprazole, atenolol, atomoxetine, bisoprolol, brexpiprazole, cavediol, clomipramine, clonidine, clozapine, codeine, desipramine, doxepin, duloxetine, eliglustat, flecainide, fluoxetine, fluphenazine, fluvoxamine, haloperidol, hydrocodone, iloperidone, imipramine, methylphenidate, metoprolol, mirtazapine, moclobemide, nebivolol, nortriptyline, odansetron, olanzapine, oxycodone, paroxetine, perphenazine, pimozone, propafenone, propranolol, quetiapine, risperidone, tamoxifen, tetrabenazine, tramadol, tropisetron, trimipramine, venlafaxine, vortioxetine, zuclopenthixol
CYP3A5	Tacrolimus
HLA-A	Carbamazepine (*31:01)
HLA-B	Abacavir (*57:01), allopurinol (*58:01), carbamazepine (*15:02), oxcarbazepine (*15:02), phenytoin (*15:02)
NUDT15	Azathioprine, mercaptopurine, thioguanine
SLCO1B1	Atorvastatin, fluvastatin, rosuvastatin, simvastatin
TPMT	Azathioprine, mercaptopurine, thioguanine
VKORC1	Acenocoumarol, warfarin

*Phenotype adjusted based on the concomitant use of inhibitors or inducers. See the Regarding Phenotype Adjustment section at the end of the report for full details.

Current Medications

Medication name	Description
esomeprazole	CYP3A5 Weak Inhibitor, Substrate CYP2C19 Strong Inhibitor, Substrate

*Note: Inhibitor and inducer information was based on the [Drug Interactions Flockhart Table](#). If a current medication is linked to a guideline supported by this tool, it will appear in the 'Future Medications' table of this report.

Pathogen detection

Pathogen detection for infectious disease

Target pathogens	CT (Chlamydia trachomatis)
	MG (Mycoplasma genitalium)
	UU (Ureaplasma urealyticum)
	UP (Ureaplasma parvum)
	NG (Neisseria gonorrhoea)
	MH (Mycoplasma hominis)
	TV (Trichomonas vaginalis)
	GV (Gardnerella Vaginalis)
	CA (Candida Albicans)
	TP (Treponema pallidum)
	HSV1
	HSV2
Internal control	hARF3

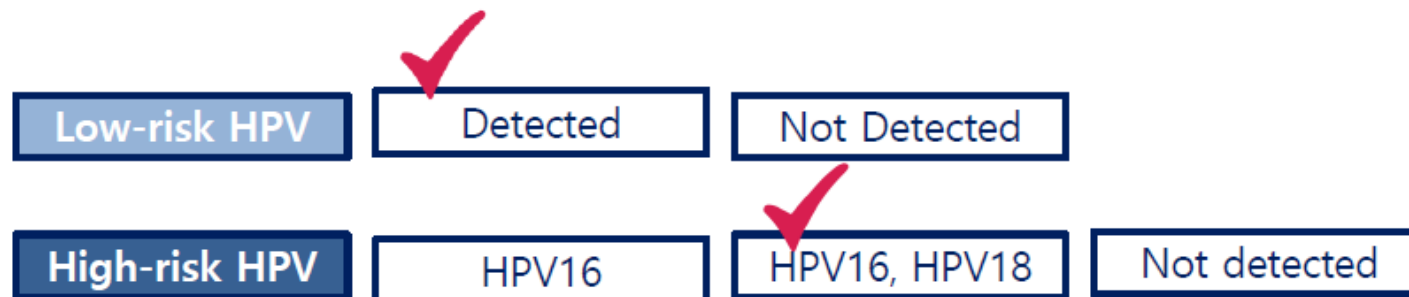
STD (Sexually transmitted disease)

HPV (Human Papilloma Virus)

- Low risk
HPV 6, 11
- High risk
HPV 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, 68

HPV (Human Papilloma Virus)

- **Targets; Low-risk HPVs, High-risk HPVs**
 - Low risk: HPV 6, 11
 - High risk: HPV 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, 68
- **Multiplexing in 1 well (total 17 targets+1 internal control)**
- **Screening program for Health care check-up purpose**



Pathogen detection

Mycobacterium strains identification & Drug-resistant tuberculosis mutation detection

W1 identified strains		W2 identified strains	
Mycobacterium tuberculosis complex		M. mucogenicum	M. lentiflavum
Mycobacterium avium complex		M. Terraе	M. marinum
M. Abcessus	M. Malmoense	M. peregrinum	M. simiae
M. intracellulare	M. Scrofulaceum	M. chimaera	M. gastri
M. asiaticum	M. smegmatis	M. gordonae	M. phlei
M. fortuitum infection	M. szulgai	M. genavense	M. septicum
M. haemophilum	M. xenopi	M. Triviale	M. massiliense
M. kansasii	M. celatum		
M. stutzeri	M. chelonae		
M. Ulcerans			

Catalog	Drug name	Resistance gene
First-line drug	Isoniazid	katG, inhA
	Rifampicin	rpoB
	Streptomycin	rpsL, rrs
	Ethambutol	embB
	Pyrazinamide	pncA
Second line drug	Fluoroquinolone	gyrA, gyrB
	Ethionamide Prothionamide	inhA
	Para-amino salicylic acid	thyA
	Clofazimine	Rv0678 (mmpR)
	amikacin kanamycin	rrs, eis
	Capreomycin	rrs
	Cycloserine	alr
	Linezolid	rplC, rrl
	rifape ntine rifab utin	rpoB

OPEN

Rapid Sputum Multiplex Detection of the *M. tuberculosis* Complex (MTBC) and Resistance Mutations for Eight Antibiotics by Nucleotide MALDI-TOF MS

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Kang-Yi Su^{1,2}, Bo-Shiun Yan³, Hao-Chieh Chiu^{1,2}, Chong-Jen Yu⁴, So-Yi Chang³, Ruwen Jou⁵, Jia-Long Liu², Po-Ren Hsueh^{2,4,*} & Sung-Liang Yu^{1,2,6,7,8,*}

The increasing incidence of multidrug-resistant (MDR) and extensively drug-resistant (XDR) *Mycobacterium tuberculosis* (MTB) adds further urgency for rapid and multiplex molecular testing to identify the MTB complex and drug susceptibility directly from sputum for disease control. A nucleotide matrix-assisted-laser-desorption-ionization time-of-flight mass spectrometry (MALDI-TOF MS)-based assay was developed to identify MTB (MTBID panel) and 45 chromosomal mutations for resistance to eight antibiotics (MTBDR panel). We conducted a 300 case trial from outpatients to evaluate this platform. An MTBID panel specifically identified MTB with as few as 10 chromosome DNA copies. The panel was 100% consistent with an acid-fast stain and culture for MTB, nontuberculous mycobacteria, and non-mycobacteria bacteria. The MTBDR panel was validated using 20 known MDR-MTB isolates. In a 64-case double-blind clinical isolates test, the sensitivity and specificity were 83% and 100%, respectively. In a 300-case raw sputum trial, the MTB identification sensitivity in smear-negative cases using MALDI-TOF MS was better than the COBAS assay (61.9% vs. 46.6%). Importantly, the failure rate of MALDI-TOF MS was better than COBAS (11.3% vs. 26.3%). To the best of our knowledge, the test described herein is the only multiplex test that predicts resistance for up to eight antibiotics with both sensitivity and flexibility.

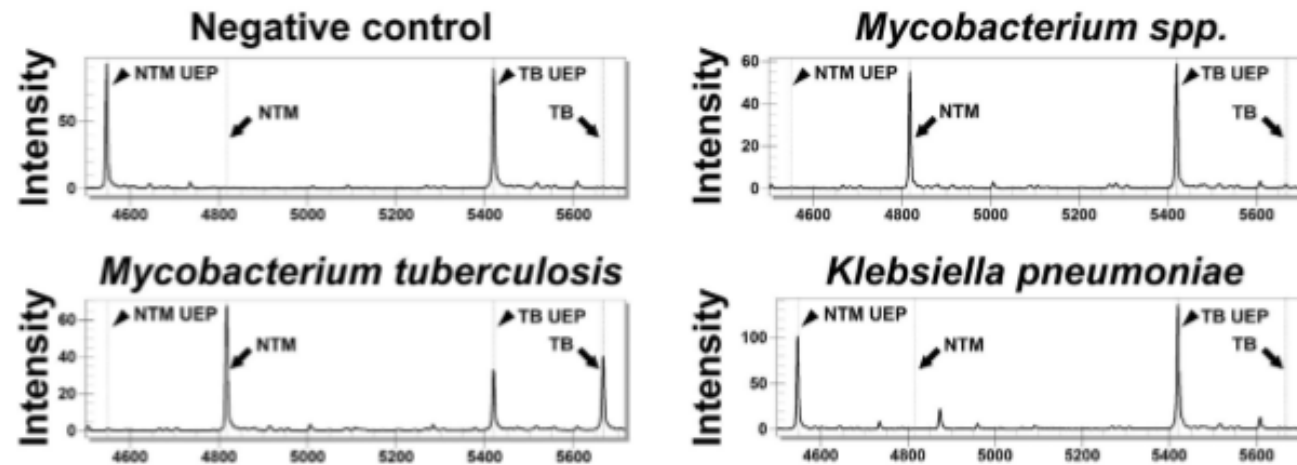
A**MCE3B**

65 75 85 95 105 115
 ACGGCAAGACCTACTACGCCGAGTTCGCCAACGTGTCCAATCTGCGAACGGCAAG
 C →

B**gyrA**

<i>M. avium</i>	CGAGTTGCCGTATCAGGTCAACCACGACAACCTTCATCACCTCGATCGCCGAGCAGGTGCGCG
<i>M. bovis</i>	CGAGTTGCCGTATCAGGTCAACCACGACAACCTTCATCACTTCGATCGCCGAACAGGTCCGAG
<i>M. tuberculosis</i>	CGAGTTGCCGTATCAGGTCAACCACGACAACCTTCATCACTTCGATCGCCGAACAGGTCCGAG
<i>M. leprae</i>	TGAGCTACCGTATCAGGTCAACCACGACAACCTTCATCACTTCTATCGCTGAGCAAGTCCGCA
<i>M. ulcerans</i>	CGAGCTGCCGTATCAGGTCAACCACGACAACCTTCATCACCTCGATCGCCGAGCAGGTTGCGG
<i>M. KMS</i>	CGAATTGCCGTATCAGGTCAACCACGACAACCTTCATCACCTCGATCGCCGAGCAGGTCCGCG
<i>M. MCS</i>	CGAATTGCCGTATCAGGTCAACCACGACAACCTTCATCACCTCGATCGCCGAGCAGGTCCGCG
<i>M. smegmatis</i>	CGAGCTGCCCTACCAGGTCAACCACGACAACCTTCATCACCTCGATCGCCGAGCAGGTGCGCG
<i>M. vanbaalenii</i>	CGAGTTGCCCTATCAGGTCAACCACGACAACCTTCATCACCTCGATCGCCGAACAGGTGCGTG

TA →

C

Publications

Design and development of MassARRAY-based bacteriological assay 10 BACTERIAL FOODBORNE PATHOGENS IN A SINGLE REACTION

Primer ID	Bacterial Targets (species)
Bac16_bac1/2	Bacteria
Eco001N	<i>E.coli/Shigella spp.</i>
Ent001	<i>Enterococcus faecalis</i>
Ent003	<i>Enterococcus faecium</i>
Clos001	<i>Clostridium perfringens</i>
Cmp002	<i>Campylobacter jejuni</i>
Cmp005	<i>Campylobacter coli</i>
Cmp006	<i>Campylobacter coli</i>
Lis001	<i>Listeria monocytogenes</i>
Stp001	<i>Staphylococcus aureus</i>
Sal002	<i>Salmonella spp.</i>

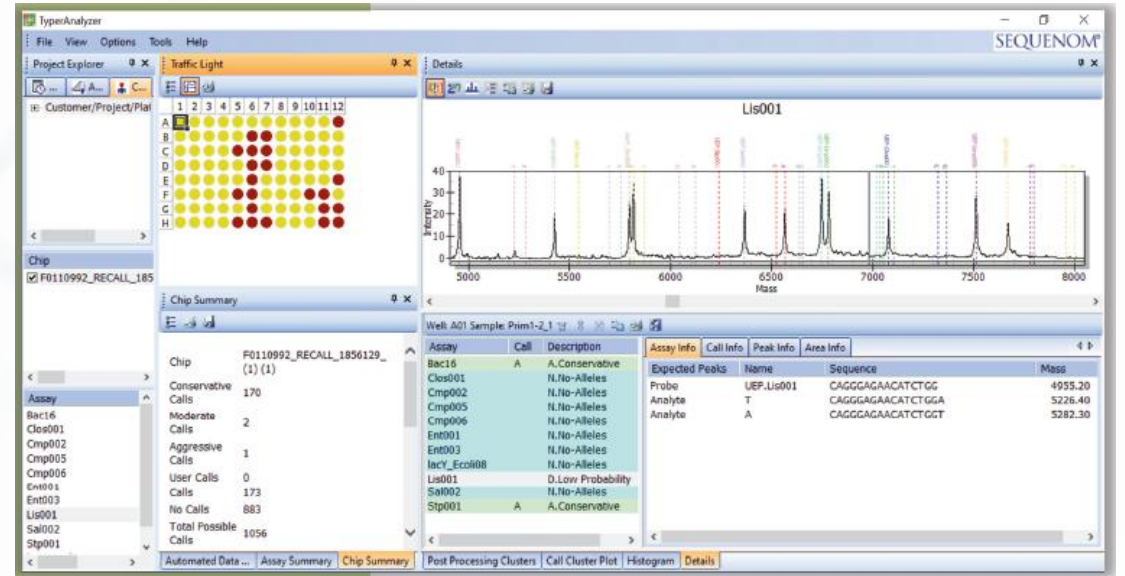


Figure 4 : TyperAnalyzer software for analysis of multiplexing reaction correlated to specific well on SpectroChip.

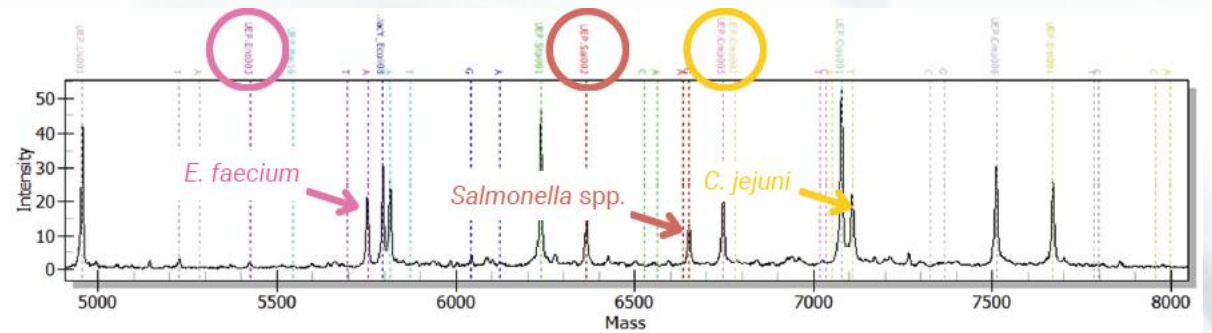
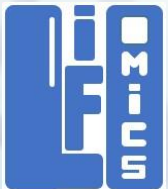


Figure 5 : *Enterococcus faecium*, *Salmonella spp.*, and *Campylobacter jejuni* were identified in a single assay. The circles represent the mass-to-charge ratio (m/z) of unextended primers (UEP), while the arrows indicate the mass spectral peak corresponding to the extended base, facilitating bacterial identification.



MassARRAY System workflow



1

2

3

4

5

Select Target
Maker

Design primer

Optimize &
validation

Sample run

Analyze data &
Report

Customized

- Assay By AGENA
- Partner
- Own panel

Disease		Wellness
SMA		Thailand : Kin Yoo Dee
Thalassemia		Nutrigenomics
Hearing loss		Exercise
TB		Korea Panel
STD		Epi-clock (Epigenetic)
Encephalitis		Cancer Disease Susceptibility
		Gut Microbiome
		Skin & Hair
		chronic disease



CUSTOMIZE

MASSCLEAVE
iPLEX Pro methylation
Methylation
Epigenetics



CUSTOMIZE

iPLEX Pro
SNP, Insertion, Deletion
Translocation
copy number variant,
somatic mutation